



**University of
Zurich**^{UZH}

**Zurich Open Repository and
Archive**

University of Zurich
University Library
Strickhofstrasse 39
CH-8057 Zurich
www.zora.uzh.ch

Year: 2016

L1-CAM is commonly expressed in testicular germ cell tumours

Fankhauser, Christian D ; Bode, Peter K ; Hermanns, Thomas ; Sander, Sophia ; Beyer, Joerg ; Sulser, Tullio ; Altevogt, Peter ; Moch, Holger ; Tischler, Verena

DOI: <https://doi.org/10.1136/jclinpath-2016-203603>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-123936>

Journal Article

Accepted Version

Originally published at:

Fankhauser, Christian D; Bode, Peter K; Hermanns, Thomas; Sander, Sophia; Beyer, Joerg; Sulser, Tullio; Altevogt, Peter; Moch, Holger; Tischler, Verena (2016). L1-CAM is commonly expressed in testicular germ cell tumours. *Journal of Clinical Pathology*, 69(5):460-462.

DOI: <https://doi.org/10.1136/jclinpath-2016-203603>

L1-CAM is commonly expressed in testicular germ cell tumors

Christian D Fankhauser^{1#*}, Peter K Bode^{2*}, Thomas Hermanns¹, Sophia Sander², Joerg Beyer³, Tullio Sulser¹, Peter Altevogt³, Holger Moch², Verena Tischler²

¹ Department of Urology, ² Institute of Surgical Pathology, ³ Department of Oncology, University Hospital of Zurich, University of Zurich, Zurich, Switzerland

⁴ Skin Cancer Unit, German Cancer Research Center (DKFZ), Heidelberg, Germany

*contributed equally

#corresponding author

Christian Daniel Fankhauser

Department of Urology, University Hospital of Zurich, University of Zurich, Frauenklinikstrasse 10, 8091 Zurich, Switzerland

Tel.: 0041442551111

Christian.fankhauser@usz.ch

Keywords

TESTIS

CANCER

UROPATHOLOGY

Word count: 974 words

Abstract

Germ cell tumors (GCT) are curable cancers, but 10-15% of patients with metastatic disease fail cisplatin-based first-line therapy. While therapeutic options have increased for various other cancers no therapeutic targets have emerged in cisplatin-refractory GCT. L1 cell adhesion molecule (L1-CAM) is commonly expressed in human malignancies and therefore a potential target, however its expression in GCT has not been studied so far. The aim of our study was to describe the expression of L1-CAM in a large series of testicular GCT. Immunohistochemistry was used to study L1-CAM expression in 325 testicular GCT, including 94 mixed GCT. L1-CAM expression was found in 38% of seminomas, 50% of yolk sac tumors, 19% of teratomas, 50% of choriocarcinomas, 67% of embryonal carcinoma. L1-CAM was expressed in 45% of germ cell neoplasias in situ but not in normal tissue. This common L1-CAM expression in testicular GCT might serve as an immunotherapeutic target in the future.

INTRODUCTION

In 2015, an estimated number of 8,430 new cases of GCT will be diagnosed in the United States [1]. Although GCTs show a high sensitivity to cisplatin-based chemotherapy, 10-15% of patients fail first-line chemotherapy and 3-5% of all GCT patients will eventually die of their disease [2]. Despite a response rate above 95% to cisplatin based chemotherapy, the search for new treatment strategies remains worthwhile in accordance to reduce treatment toxicity and offer therapeutic options in non-responding patients [3-5]. Various tumors have been described to express L1-CAM including lung carcinoma, gliomas, melanoma, renal, ovarian, endometrial and colon carcinoma [6]. L1-CAM is associated with tumor cell dissemination via the regulation of pro-metastatic MMP-2 and MMP-9 in solid and non-solid tumors [7] as well as in brain metastases[8]. L1-CAM is involved in epithelial to mesenchymal transition (EMT)[9]. In various malignancies there is evidence showing that expression of L1-CAM is associated with a subset of highly aggressive tumors with adverse clinical outcome and might serve as a therapeutic target[10]. We aimed to investigate the expression of L1-CAM in the different GCT subtypes.

MATERIALS AND METHODS

The construction of the tissue micro array (TMA) was described before [9]. L1-CAM immunohistochemistry (IHC) was performed using the monoclonal antibody anti-L1-CAM (clone 14.10, directed to the ecto-domain, 1:200). The antibody was generated as described previously [11] and was tested on a multi tissue TMA for the appropriate dilution. Peripheral neuronal tissue served as internal positive control for L1-CAM staining. Sertoli cells and Leydig cells were negative for L1-Cam. Two experienced surgical pathologists (PKB, VT) evaluated the L1-CAM stained TMA. Samples were dichotomized into positive versus negative. The threshold for positivity was defined at 5% of cells with a moderate or strong staining. The expression patterns were separately analyzed for each tumor component, GCNIS and normal tissue. The study was approved by the local ethics committee (reference number KEK StV. 25-2008).

RESULTS

The series included a total of 207 seminomas, 4 spermatocytic tumours, 19 pure embryonal carcinomas, 1 pure mature teratoma and 94 mixed GCT. Mixed germ cell tumors included the following components: seminoma, embryonal carcinoma, yolk sac tumor, choriocarcinoma and teratoma (49 with and 45 without a seminomatous component).

The individual tumor components consisted of 253 seminomas, 89 embryonal carcinomas, 52 yolk sac tumors, 53 teratomas, 10 choriocarcinomas. In addition, we included non-tumorous testicular tissue from 20 tumor patients and GCNIS from 20 patients.

L1-CAM IHC staining showed an exclusively membranous staining pattern in GCTs with moderate to strong intensity. In seminoma, IHC staining of L1-CAM showed a predominantly heterogeneous membranous pattern and 95 cases (38%) were L1-CAM positive (see table and figure 1).

Tissue types	Negative	Positive	Positive cases in %
Normal testis (n=20)	20	0	0%
Intratubular germ cell neoplasia (n=20)	11	9	45%

Seminoma (n=253)	158	95	38%
Choriocarcinoma (n=10)	5	5	50%
Yolk sack tumor (n=52)	26	26	50%
Embryonal carcinoma (n=89)	29	60	67%
Teratoma (n=53)	43	10	19%

Table 1 L1-CAM expression in non-neoplastic and neoplastic testicular tissue

The yolk sac tumors with microcystic, glandular, solid and spindle cell growth patterns showed a heterogeneous expression of L1-CAM in 26 cases (50%). The glandular components were positive in a L1-CAM staining whereas areas with stromal or spindle cell differentiation were negative. The results of L1-CAM staining in teratoma components were very heterogeneous depending on the tissue types found in the teratoma. Ten cases (19%) showed L1-CAM positive structures, e.g. glands with a membranous pattern. Stromal components as well as chondrocytes were L1-CAM negative. Five (50%) choriocarcinoma cases expressed L1-CAM. The expression was more restricted to the syncytiotrophoblastic giant cells whereas the mononuclear component was predominantly negative. Interestingly, syncytiotrophoblastic giant cells were also positive when scattered in a seminomatous tissue component. In embryonal carcinoma 59 cases (63%) were positive with an intense and homogenous pattern. Normal testicular tissue was L1-CAM negative whereas IGCNU showed a strong expression of L1-CAM in 45% of all cases.

DISCUSSION

Recently, L1-CAM has emerged as a potential therapeutic target due to its expression on many solid tumors, and only limited expression on normal tissues[6]. *In-vitro* and *in-vivo* studies showed efficacy and safety of L1-CAM targeting antibodies by acting via antibody-dependent cellular cytotoxicity (ADCC) or by being labeled with radionuclides[12-23]. In a humanized transgenic mouse model of L1-CAM no adverse effects were observed after injection of ant-L1-CAM antibodies [24]. A first in human phase I trial was

published by Park et al showed no adverse effects and some objective response after infusing autologous CE7R/HyTK+ CD8+ cytolytic T-lymphocytes [20]. However, it cannot be concluded from this study, if the response was due to the L1-CAM therapy or the subsequent salvage therapies.

We observed that L1-CAM expression is markedly enhanced in most GCTs, in 45% of GCNIS but not in normal tissue. As typical for a retrospective study, this investigation is limited by potential biases, such as patient selection and bias of core punching.

To our knowledge this is the first publication describing L1-CAM expression in GCTs. We conclude that the frequent expression of L1-CAM in testicular seminomas and non-seminomas indicates that L1-CAM could be a promising new therapeutic target in testicular cancer that warrants further functional studies and potentially investigation in clinical trials in the future. Because L1-CAM was never expressed on normal but frequently on cancer cells, further investigation should elaborate the role for L1-CAM as a new "neoplastic" germ cell tumor marker.

Acknowledgments: We thank Martina Storz and Susanne Dettwiler for excellent technical assistance, and Peter Schraml, Tumor Tissue Bank, University Hospital Zurich, for help with tissue microarray construction.

Competing Interests: None declared.

Funding: None

Licence for Publication: The Corresponding Author has the right to grant on behalf of all authors and does grant on behalf of all authors, an exclusive licence (or non exclusive for government employees) on a worldwide basis to the BMJ Publishing Group Ltd to permit this article (if accepted) to be published in JCP and any other BMJ PGL products and sublicences such use and exploit all subsidiary rights, as set out in our licence (<http://group.bmj.com/products/journals/instructions-for-authors/licence-forms>).

Figure Labelling

Figure 1. L1 cell adhesion molecule (L1-CAM)staining in several tumor components: Classic seminoma with heterogeneous membranous (A); Embryonal carcinoma with typical homogenous and intense membranous staining pattern (B); Yolk sac tumor with weak heterogeneous membranous staining (C); choriocarcinoma with strong membrane staining in dispersed tumor cells, predominantly multinucleated giant cells (D); teratoma with heterogeneous membranous staining (E); strong staining in the basal part of the tubuli in carcinoma-in-situ (CIS) germ cells (F)

References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. *CA Cancer J Clin* 2015;**65**(1):5-29 doi: 10.3322/caac.21254[published Online First: Epub Date]].
2. Gandaglia G, Becker A, Trinh Q-D, et al. Long-term survival in patients with germ cell testicular cancer: a population-based competing-risks regression analysis. *Eur J Surg Oncol* 2014;**40**(1):103-12 doi: 10.1016/j.ejso.2013.09.019[published Online First: Epub Date]].
3. Fankhauser CD, Honecker F, Beyer J, Bode PK. Emerging Therapeutic Targets for Male Germ Cell Tumors. *Curr Oncol Rep* 2015;**17**(12):54 doi: 10.1007/s11912-015-0479-4[published Online First: Epub Date]].
4. Travis LB, Fossa SD, Schonfeld SJ, et al. Second cancers among 40,576 testicular cancer patients: focus on long-term survivors. *Journal of the National Cancer Institute* 2005;**97**(18):1354-65 doi: 10.1093/jnci/dji278[published Online First: Epub Date]].
5. Jacobsen C, Honecker F. Cisplatin resistance in germ cell tumours: models and mechanisms. *Andrology* 2015;**3**(1):111-21 doi: 10.1111/andr.299[published Online First: Epub Date]].
6. Altevoigt P, Doberstein K, Fogel M. L1CAM in human cancer. *International journal of cancer. Journal international du cancer* 2015 doi: 10.1002/ijc.29658[published Online First: Epub Date]].
7. Weinspach D, Seubert B, Schaten S, et al. Role of L1 cell adhesion molecule (L1CAM) in the metastatic cascade: Promotion of dissemination, colonization, and metastatic growth. *Clinical and Experimental Metastasis* 2014;**31**(1):87-100
8. Valiente M, Obenauf AC, Jin X, et al. Serpins promote cancer cell survival and vascular co-option in brain metastasis. *Cell* 2014;**156**(5):1002-16 doi: 10.1016/j.cell.2014.01.040[published Online First: Epub Date]].
9. Tischler V, Pfeifer M, Hausladen S, et al. L1CAM protein expression is associated with poor prognosis in non-small cell lung cancer. *Molecular cancer* 2011;**10**:127 doi: 10.1186/1476-4598-10-127[published Online First: Epub Date]].
10. Gavert N, Ben-Shmuel A, Raveh S, Ben-Ze'ev A. L1-CAM in cancerous tissues. *Expert opinion on biological therapy* 2008;**8**(11):1749-57 doi: 10.1517/14712598.8.11.1749[published Online First: Epub Date]].
11. Huszar M, Moldenhauer G, Gschwend V, Ben-Arie A, Altevoigt P, Fogel M. Expression profile analysis in multiple human tumors identifies L1 (CD171) as a molecular marker for differential diagnosis and targeted therapy. *Human pathology* 2006;**37**(8):1000-8 doi: 10.1016/j.humpath.2006.03.014[published Online First: Epub Date]].

12. Arlt MJ, Novak-Hofer I, Gast D, et al. Efficient inhibition of intra-peritoneal tumor growth and dissemination of human ovarian carcinoma cells in nude mice by anti-L1-cell adhesion molecule monoclonal antibody treatment. *Cancer research* 2006;**66**(2):936-43 doi: 10.1158/0008-5472.can-05-1818[published Online First: Epub Date]].
13. Fischer E, Grunberg J, Cohrs S, et al. L1-CAM-targeted antibody therapy and (177)Lu-radioimmunotherapy of disseminated ovarian cancer. *International journal of cancer. Journal international du cancer* 2012;**130**(11):2715-21 doi: 10.1002/ijc.26321[published Online First: Epub Date]].
14. Gast D, Riedle S, Issa Y, et al. The cytoplasmic part of L1-CAM controls growth and gene expression in human tumors that is reversed by therapeutic antibodies. *Oncogene* 2008;**27**(9):1281-9 doi: 10.1038/sj.onc.1210747[published Online First: Epub Date]].
15. Grünberg J, Lindenblatt D, Dorrer H, et al. Anti-L1CAM radioimmunotherapy is more effective with the radiolanthanide terbium-161 compared to lutetium-177 in an ovarian cancer model. *European journal of nuclear medicine and molecular imaging* 2014;**41**(10):1907-15 doi: 10.1007/s00259-014-2798-3[published Online First: Epub Date]].
16. Hoefnagel CA, Rutgers M, Buitenhuis CK, et al. A comparison of targeting of neuroblastoma with mIBG and anti L1-CAM antibody mAb chCE7: therapeutic efficacy in a neuroblastoma xenograft model and imaging of neuroblastoma patients. *European journal of nuclear medicine* 2001;**28**(3):359-68
17. Knogler K, Grünberg J, Zimmermann K, et al. Copper-67 radioimmunotherapy and growth inhibition by anti-L1-cell adhesion molecule monoclonal antibodies in a therapy model of ovarian cancer metastasis. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2007;**13**(2 Pt 1):603-11 doi: 10.1158/1078-0432.ccr-06-1486[published Online First: Epub Date]].
18. Lindenblatt D, Fischer E, Cohrs S, Schibli R, Grunberg J. Paclitaxel improved anti-L1CAM lutetium-177 radioimmunotherapy in an ovarian cancer xenograft model. *EJNMMI Research* 2014;**4**(1):54
19. Novak-Hofer I, Cohrs S, Grunberg J, et al. Antibodies directed against L1-CAM synergize with Genistein in inhibiting growth and survival pathways in SKOV3ip human ovarian cancer cells. *Cancer letters* 2008;**261**(2):193-204 doi: 10.1016/j.canlet.2007.11.012[published Online First: Epub Date]].
20. Park JR, Digiusto DL, Slovak M, et al. Adoptive transfer of chimeric antigen receptor re-directed cytolytic T lymphocyte clones in patients with neuroblastoma. *Molecular therapy : the journal of the American Society of Gene Therapy* 2007;**15**(4):825-33 doi: 10.1038/sj.mt.6300104[published Online First: Epub Date]].
21. Primiano T, Baig M, Maliyekkel A, et al. Identification of potential anticancer drug targets through the selection of growth-inhibitory genetic suppressor elements. *Cancer cell* 2003;**4**(1):41-53
22. Schafer H, Dieckmann C, Kornienko O, et al. Combined treatment of L1CAM antibodies and cytostatic drugs improve the therapeutic response of pancreatic and ovarian carcinoma. *Cancer letters* 2012;**319**(1):66-82 doi: 10.1016/j.canlet.2011.12.035[published Online First: Epub Date]].
23. Wolterink S, Moldenhauer G, Fogel M, et al. Therapeutic antibodies to human L1CAM: functional characterization and application in a mouse model for ovarian carcinoma. *Cancer research* 2010;**70**(6):2504-15 doi: 10.1158/0008-5472.can-09-3730[published Online First: Epub Date]].
24. Doberstein K, Harter PN, Haberkorn U, et al. Antibody therapy to human L1CAM in a transgenic mouse model blocks local tumor growth but induces EMT. *International journal of cancer. Journal international du cancer* 2015;**136**(5):E326-39 doi: 10.1002/ijc.29222[published Online First: Epub Date]].